

**UCC Library and UCC researchers have made this item openly available.  
 Please [let us know](#) how this has helped you. Thanks!**

<b>Title</b>	Sphingosine 1-phosphate, a potential target in neovascular retinal disease
<b>Author(s)</b>	Alshaikh, Rasha A.; Ryan, Katie B.; Waeber, Christian
<b>Publication date</b>	2021-05-07
<b>Original citation</b>	Alshaikh, R. A., Ryan, K. B. and Waeber, C. (2021) 'Sphingosine 1-phosphate, a potential target in neovascular retinal disease', British Journal of Ophthalmology. doi: 10.1136/bjophthalmol-2021-319115
<b>Type of publication</b>	Article (peer-reviewed)
<b>Link to publisher's version</b>	<a href="http://dx.doi.org/10.1136/bjophthalmol-2021-319115">http://dx.doi.org/10.1136/bjophthalmol-2021-319115</a> Access to the full text of the published version may require a subscription.
<b>Rights</b>	© 2021, the Authors. Published by BMJ Publishing Group Ltd. This article has been accepted for publication in British Journal of Ophthalmology following peer review, and the Version of Record can be accessed online at: <a href="http://dx.doi.org/10.1136/bjophthalmol-2021-319115">http://dx.doi.org/10.1136/bjophthalmol-2021-319115</a> No commercial re-use. <a href="https://creativecommons.org/licenses/by-nc/4.0/">https://creativecommons.org/licenses/by-nc/4.0/</a>
<b>Item downloaded from</b>	<a href="http://hdl.handle.net/10468/11391">http://hdl.handle.net/10468/11391</a>

Downloaded on 2021-11-27T16:47:21Z

# **Sphingosine 1-phosphate, a potential target in neovascular retinal disease**

Rasha A. Alshaikh <sup>a,b</sup>, Katie B. Ryan <sup>a,c</sup>, Christian Waeber <sup>a,d,\*</sup>

<sup>a</sup> School of Pharmacy, University College Cork, Cork, Ireland

<sup>b</sup> Department of Pharmaceutical Technology, Faculty of Pharmacy, Tanta University, Tanta, Egypt.

<sup>c</sup> SSPC The SFI Research Centre for Pharmaceuticals, School of Pharmacy, University College Cork

<sup>d</sup> Department of Pharmacology and Therapeutics, University College Cork, Cork, Ireland

\* Corresponding author at: School of Pharmacy, University College Cork, College Rd, Cork T12 YN60, Ireland.

E-mail address: [c.waeber@ucc.ie](mailto:c.waeber@ucc.ie)

## **ABSTRACT**

Neovascular ocular diseases (such as age-related macular degeneration, diabetic retinopathy and retinal vein occlusion) are characterized by common pathological processes that contribute to disease progression. These include angiogenesis, edema, inflammation, cell death and fibrosis. Currently available therapies target the effects of vascular endothelial growth factor (VEGF), the main mediator of pathological angiogenesis. Unfortunately, VEGF blockers are expensive biological therapeutics that necessitate frequent intravitreal administration and are associated with multiple adverse effects. Thus, alternative treatment options associated with lower side effects are required for disease management. This review introduces sphingosine 1-phosphate (S1P) as a potential pharmacological target for treatment of neovascular ocular pathologies. S1P is a sphingolipid mediator that controls cellular growth, differentiation, survival and death. S1P actions are mediated by five G Protein Coupled Receptors (S1P<sub>1-5</sub> receptors) which are abundantly expressed in all retinal and sub-retinal structures. The action of S1P on S1P<sub>1</sub> receptors can reduce angiogenesis, increase endothelium integrity, reduce photoreceptor apoptosis and protect the retina against neurodegeneration. Conversely, S1P<sub>2</sub> receptor signaling can increase neovascularization, disrupt endothelial junctions, stimulate VEGF release, induce retinal cell apoptosis and degeneration of neural retina. The aim of this review is to thoroughly discuss the role of S1P and its different receptor subtypes in angiogenesis, inflammation, apoptosis and fibrosis in order to determine which of these S1P-mediated processes may be targeted therapeutically.

## **Key words**

Sphingosine-1 phosphate, Neovascularization, VEGF blockers, Angiogenesis, Age-related macular degeneration, Diabetic retinopathy, Retinal vein occlusion.

## INTRODUCTION

At least 2.2 billion people suffer from vision impairment, which in 1 billion cases can be attributed to a preventable or treatable cause. Diabetic retinopathy and age-related macular degeneration account for more than one third of these cases.[1] Neovascular ocular disease including diabetic retinopathy (DR), wet age-related macular degeneration (wet-AMD) and retinal vein occlusion (RVO) have different etiology but result in a similar cascade of pathophysiological events (Table 1). Pathological angiogenesis is key amongst these, and a hallmark of disorders that occurs in response to vascular endothelial growth factor (VEGF), a potent hypoxia-induced angiogenic mediator that triggers the formation of new permeable and unstable blood vessels.[2] Pathological angiogenesis can originate from retinal vasculature which supplies the inner retina and/or choroidal vasculature which supplies the outer retinal and retinal pigment epithelium (RPE). RPE is a monolayer of epithelial cells that represent the main structure of outer blood retinal barrier,[3,4] and prevents retinal invasion of neovascular tissue of choroidal blood vessels.[5] In neovascular ocular disease, angiogenesis is accompanied with disruption of RPE physical and metabolic barrier function (Table 1). This results in continuous leakage of blood or blood components to the surrounding tissues, leading to edema and/or hemorrhage, with possible progression to retinal detachment and irreversible apoptosis of photoreceptors and other retinal cells (Table 1). In addition to angiogenesis, chronic hypoxia causes chronic inflammation and over-production of reactive oxygen species in the retina, triggering cell death and fibrotic cascades (Table 1). Fibrosis in the posterior chamber of the eye has unique characteristics, being characterized by the occurrence of gliosis and epithelial to mesenchymal transition (EMT). Later sections describe the role of S1P in both gliosis and EMT.

**Table 1.** Common pathological events associated with different neovascular ocular diseases. Most of these have been shown to be affected by S1P signaling.

Pathological event	Occurrence in different neovascular ocular disease					
	AMD	DR	RVO	RP	ROP	Glaucoma
Retinal and subretinal neovascularization	[6,7]	[7,8]	[7,9]			
Macular edema	[10]	[7,10]	[7,10]			
Disruption of RPE barrier function		[11]				
Apoptosis or degeneration of photoreceptors, RPE and other retinal cells	[12,13]	[14,15]		[16]		
Chronic inflammation	[17]	[18]		[19]	[20]	
Ganglionic cell death and retinal neurodegeneration	[21]	[22]				[23,24]
Retinal and extraretinal fibrosis	[25]	[26]				

AMD is age related macular degeneration, DR is diabetic retinopathy, RVO is retinal vein occlusion, ROP is retinopathy of prematurity and RPE is retinal pigmented epithelium.

Angiogenesis, edema, inflammation, apoptosis and fibrosis contribute to the pathophysiology of DR, wet-AMD and RVO (Table 1). Current therapeutic options largely rely on the blockade of VEGF signaling with biological therapeutics (antibodies, recombinant fusion proteins or pegylated RNA aptamers). They are expensive, with limited stability and are administered by invasive intravitreal injections that can be associated with retinal detachment, subconjunctival hemorrhage, uveitis, and endophthalmitis.[27] It would therefore be beneficial to develop novel therapeutic agents with fewer limitations. The lipid mediator sphingosine 1-phosphate (S1P) is involved in hypoxia-induced angiogenesis. Unlike VEGF, S1P can promote stable blood vessels formation, increase endothelial barrier integrity and positively impact the subsequent pathophysiological steps leading to neovascular ocular diseases.

This review will provide a brief description of S1P metabolism, the distribution and function of S1P receptor subtypes in different retinal tissues, and will then more specifically focus on the documented effects of S1P and the sometimes opposing roles of various S1P receptor subtypes in the processes that contribute to neovascular disease pathogenesis.

## **S1P PRODUCTION, METABOLISM AND RECEPTOR EXPRESSION IN OCULAR TISSUES**

Sphingolipids are lipid-based cell membrane components. In addition to their structural role, they modulate cellular proliferation, migration, differentiation, and survival.[28,29] They are synthesized by serine palmitoyl transferase (SPT) from palmitoyl CoA and serine as summarized in Figure 1. S1P is produced by phosphorylation of sphingosine by one of two sphingosine kinase isoforms, SphK1 and SphK2.[30] S1P can be de-phosphorylated by the action of two phosphatases, or degraded by S1P lyase to produce inactive metabolites (Figure 1).[31]

The regulation of S1P production and release in different body tissues is not yet completely understood. In plasma, S1P is mainly produced by red blood cells (RBCs), endothelial cells, and platelets. Once produced intracellularly, S1P is transported to the extracellular space leading to significantly higher plasma concentrations of the mediator ( $\sim 1 \mu\text{M}$ ) compared to interstitial fluid levels.[32] Most circulating S1P is not free, but bound to high-density lipoproteins (HDL), albumin, and to lower extent low-density lipoproteins (LDL).

Under normal conditions, SphK2 is the main S1P-producing kinase in rat and mouse retina.[33,34] Under hypoxic or light-induced stress conditions, SphK1 but not SphK2 is upregulated leading to increased intracellular S1P levels in murine retina [33,35]. Little is known about the levels and role of potential S1P carriers in ocular tissues. Albumin can be found in fetal vitreous, the retina and lens.[36] LDL and HDL can be synthesized locally in the retina [37] or diffuse from the systemic circulation through RPE, although HDL diffusion was significantly lower than LDL in rat retina [38] Apolipoprotein E is synthesized by Müller glial cells in neural retina and transported to vitreous humor,[39] with no information describing retinal expression of apolipoprotein A4 or M.

While S1P can act as an intracellular second messenger and as an extracellular mediator, the latter effects, mediated by five G-protein coupled receptors (S1P<sub>1-5</sub>) predominate.[35] S1P<sub>1-3</sub> receptors are expressed in almost every body tissue, while S1P<sub>4</sub> and S1P<sub>5</sub> expression is largely restricted to the lymphatic and nervous systems.[40,41] S1P receptor expression in retina varies depending on cell type and pathophysiological status (see Table 2 for a summary). Under healthy conditions, S1P<sub>1</sub> receptors predominate in retina, while photoreceptors mainly express S1P<sub>2</sub> receptors (Table 2).[33,35] Retinal pigmented epithelium (RPE) cells show robust S1P<sub>1-3</sub> receptor expression, with different subtypes predominating in different cell lines.[42,43] Retinal

vasculature endothelial cells, isolated from human donor tissues predominantly express S1P<sub>2</sub> and S1P<sub>3</sub> receptors (Table 2).[44] New single cell RNA sequencing (scRNA-seq) data show that S1P<sub>1</sub> and S1P<sub>3</sub> genes are the most strongly expressed in retinal endothelial cells,[45] while choriocapillaris endothelial cells mostly express S1P<sub>1</sub>, followed by S1P<sub>3</sub>. [46] Muller glial cells express S1P<sub>1</sub> and S1P<sub>3</sub> receptors, with S1P<sub>3</sub> receptor expression most evident in peripheral rather than foveal Müller glial cells.[47] S1P receptors densities are altered under pathological conditions. S1P<sub>2</sub> and S1P<sub>3</sub>, but not S1P<sub>1</sub> receptors, are upregulated following light-induced retinal damage.[33] Likewise, a 5-fold increase in the retinal S1P<sub>2</sub> receptors expression is seen in response to hypoxia.[48] Recent reports also highlight the role of S1P<sub>2</sub> receptors in laser-induced choroidal neovascular lesions (CNV).[49] Additionally, scRNA-seq reveal noticeable downregulation of both S1P<sub>1</sub> and S1P<sub>3</sub> receptor expression in the choroidal endothelial cells pooled from an AMD patient.[46]

Growing evidence suggests that S1P plays a significant role in normal retinal development. S1P<sub>1-3</sub> receptors loss leads to significant defects in the retinal vascular network of post-natal mice.[50] Additionally, S1P has essential functions in photoreceptor development, proliferation, differentiation and survival.[51] A link between sphingolipids and ocular disease was first suspected following the observation of ocular abnormalities in sphingolipidoses, a group of lysosomal storage disorders characterized by build-up of certain sphingolipids in which retinal degeneration, neovascularization and blindness are common manifestations.[52,53] Hereditary and sensory autonomic neuropathy type 1 (HSAN1) is characterized by a mutated SPT. The defective enzyme synthesizes deoxysphingolipids lacking the hydroxyl group at C1, which is essential for synthesis of other sphingolipids. Deoxysphingolipids are cytotoxic, suggesting the involvement of these mediators in the neuropathy of HSAN1.[54] Deoxysphingolipids are also



seen in macular telangiectasia. Patients with this condition have normal SPT, but significantly lower serum serine levels, resulting in the use of alanine instead of serine as a substrate for SPT, ending in formation of deoxysphingolipids. Deoxysphingolipid serum levels in these patients is positively correlated to the disease severity.[55,56] However, a role for S1P and its receptors in these diseases has not been reported and they will not be discussed further in this review.

**Table 2.** S1P receptors expression in different tissues of the ocular posterior segment.

Posterior segment structure	Experimental condition (cell line/ animal used)	Dominant receptor expression	Other receptors expressed	Detection method	References
Retina	Adult Sprague Dawley rat eye tissue	S1P <sub>1</sub>	- S1P <sub>3</sub> has the highest expression after S1P <sub>1</sub> - S1P <sub>2</sub> , and S1P <sub>5</sub> had minimal expression	qRT-PCR	[33]
	Post-natal mice eye tissue	S1P <sub>1</sub>	- S1P <sub>3</sub> has the highest expression after S1P <sub>1</sub> -S1P <sub>2</sub> had minimal expression	qRT-PCR	[33]
Photoreceptors	661W photoreceptor cell line	S1P <sub>2</sub>	- S1P <sub>1</sub> and S1P <sub>3</sub> were detected in significantly lower levels - S1P <sub>4</sub> and S1P <sub>5</sub> were detected in negligible concentrations	qPCR	[35]
RPE	ARPE-19 cell line	S1P <sub>2</sub>	- S1P <sub>5</sub> and S1P <sub>1</sub> exhibit higher expression after S1P <sub>2</sub>	qPCR	[57]

			- S1P <sub>3</sub> was barely detected while S1P <sub>4</sub> was not detected.		
	ARPE-19 cell line	S1P <sub>3</sub>	- S1P <sub>1</sub> , S1P <sub>2</sub> , S1P <sub>4</sub> and S1P <sub>5</sub> were all expressed at lower levels.	Semi quantitative RT-PCR	[43]
	Primary human RPE	S1P <sub>1</sub> , S1P <sub>2</sub> , S1P <sub>3</sub>	- S1P <sub>4</sub> almost undetectable - S1P <sub>5</sub> has minimal expression	RT-PCR	[42]
RPE-choroid	Adult Sprague Dawley rats	S1P <sub>3</sub>	-	qRT-PCR	[33]
Retinal vasculature endothelial cells	Primary Human Retinal Endothelial Cells (HREC) isolated from donor tissue	S1P <sub>2</sub>	- S1P <sub>3</sub> was expressed in slightly lower but comparable levels to S1P <sub>2</sub> . - S1P <sub>1</sub> was expressed in significantly lower levels. - S1P <sub>4</sub> and S1P <sub>5</sub> were not significantly expressed	qRT-PCR	[44]

---

	Post-natal		- S1P <sub>1</sub> has the highest		
			expression after S1P <sub>3</sub> .		
Optic nerve	mice eye	S1P <sub>3</sub>		qRT-PCR	[33]
	tissue		- S1P <sub>2</sub> was expressed in		
			minute amounts.		

---

qRT-PCR is quantitative reverse transcriptase polymerase chain reaction, qPCR is quantitative real-time polymerase chain reaction and RT-PCR is reverse transcriptase polymerase chain reaction

The altered expression and differential effects of various S1P receptor subtypes under stress conditions offers unique opportunities to target these receptors with selective agonists or antagonists in ocular disorders. The following sections describe the role of individual S1P receptor subtypes in the relevant pathological processes. In this context, it is worth bearing in mind the uncertain specificity of the pharmacological agents used in the studies described in this review.[58] Better characterized clinical candidates, or clinically used S1P receptor drugs have been developed primarily for the management of multiple sclerosis (MS).[59,60] These include approved S1P receptors modulators fingolimod (FTY720), siponimod (BAF312) and ozanimod (RPC1063). Fingolimod binds to all S1P receptors except S1P<sub>2</sub>, siponimod and ozanimod are selective S1P<sub>1</sub> and S1P<sub>5</sub> modulators.[61] To the best of our knowledge, these agents have not been used to characterize the role of S1P signaling in ocular pathophysiology. Ocular pathologies and macular edema are associated with fingolimod therapy in MS patients,[62] but may result from MS progression rather than fingolimod treatment.[63]

## ANGIOGENESIS

Angiogenesis is the process by which new blood vessels develop from pre-existing ones. The process is carefully regulated by a balance of stimulatory (e.g., VEGF, hypoxia inducible factor (HIF), transforming growth factor- $\beta$  (TGF- $\beta$ )) and inhibitory signaling, which maintains a minimal turnover of endothelial cells. In adults, physiological angiogenesis is transient and only occurs during menstrual cycle and wound healing.[64] Under specific conditions (e.g., hypoxia, inflammation, acidosis), quiescent endothelial cells show massive proliferation and migration in a phenomenon called “the angiogenic switch”, where the influence of angiogenesis activators exceeds that of the inhibitors.[64]. The vessel formation is then initiated by the release of pro-angiogenic mediators and growth factors.[65] These factors trigger transcriptional responses in the endothelium, with specialized endothelial cells becoming “tip cells” which guide vessel branching toward the angiogenic stimulus. This is followed by enzymatic (e.g., metalloproteases) lysis of the basement membrane and extracellular matrix, while other endothelial cells proliferate and follow the lead of the tip cells. These cells differentiate into stalk cells responsible for lumen formation, basement membrane deposition, growth and quiescence of new endothelial cell and expression of intercellular junction proteins. This is accompanied by migration of pericytes and vascular smooth muscle cells to support the young blood vessels. Once blood is flowing in the new vessel, fluid shear forces act as an inhibitor of angiogenesis, a process in which S1P<sub>1</sub> receptors play a key role.[66]

Under pathological conditions (e.g., diabetic retinopathy), persistent tissue hypoxia results in sustained release of angiogenic stimulators and exhaustive endothelium activation.[67] This triggers erosion of basement membrane in multiple locations, substantial tip cell formation and continuous endothelial proliferation and migration. Due to the high concentration of angiogenic mediators, endothelial activation occurs even in newly formed vessels. This leads to fragile

endothelium with no time for maturation and subsequently no ability to restore blood flow,[64] further exacerbating hypoxia, leakage of blood components to the surrounding tissues and release of more angiogenic mediators in a vicious cycle that contributes to disease progression in many pathologies including neovascular ocular disease.[68]

## **ROLE OF S1P IN RETINAL AND CHOROIDAL ANGIOGENESIS AND BLOOD VESSEL INTEGRITY**

While S1P can be described as a hypoxia-induced pro-angiogenic mediator, this oversimplifies its complex role in vascular growth and stability.[69,70] The effect of S1P in angiogenesis is mediated mainly via S1P<sub>1</sub> and S1P<sub>2</sub> receptors, which are both highly expressed in retinal endothelium. The number and integrity of the vessels formed in response to this lipid depend on which receptor subtype is principally involved. To add further complexity, the nature of the endothelial response to S1P depends on which carrier protein (albumin, apolipoprotein A4, or M) presents the lipid to its target receptor.[71] Similar observations were made in RPE cells.[49] Finally, it is important to note that S1P<sub>1</sub> receptors are an essential component of fluid shear stress sensing and can be activated in the absence of S1P.[66]

### **S1P<sub>1</sub> receptors**

S1P<sub>1</sub> receptors trigger a distinctive and controlled angiogenic pattern where only a limited number of blood vessel sprouts are formed.[66,72–74] These sprouts undergo full maturation and acquire the characteristic of mature endothelium forming a competent vascular network.[66,75,76] S1P-mediated enhancement of endothelial integrity is abolished in S1P<sub>1</sub>-knockdown endothelium,[76] while S1P<sub>1</sub> gene deletion increases tip cell formation in the retinal vasculature, leading to hyper-sprouting, which is associated with disrupted intercellular junctions, reduced capillary perfusion and hypoxia in surrounding retinal tissues, with VEGF over-expression in

retinal endothelium.[66] Similar results are obtained in postnatal mice retinas by administering the S1P<sub>1</sub> antagonist W146.[74] Conversely, S1P<sub>1</sub> receptor overexpression reduces the number of tip cells and vessel branch points in retinal vasculature of mice embryos,[66] while S1P<sub>1</sub> receptor activation with the S1P<sub>1</sub> agonist SEW2871 results in angiogenesis characterized by fewer but longer blood vessel branches and reduces the VEGF angiogenic effect on human umbilical vein endothelial cells (HUVEC) or mouse microvascular endothelial cells.[74] As pathological neovascularization is characterized by higher VEGF levels, the ability of S1P<sub>1</sub> receptor activation to antagonize VEGF angiogenic effect is of pathological relevance.[74] Finally, the levels of cell junction proteins are reduced following S1pr1 silencing by siRNA, targeted S1pr1 gene deletion in endothelial cells, or VEGF stimulation.[74]

In conclusion, while S1P<sub>1</sub> receptor stimulation is generally pro-angiogenic, it results in the formation of fewer blood vessel sprouts, that develop competent endothelium. This can restore blood flow to the hypoxic retina without resulting in edema or hemorrhage (Figure 2).

## **S1P<sub>2</sub> receptors**

S1P<sub>2</sub> receptors activate an angiogenetic response similar to that of VEGF,[77] in which continuous, uncontrolled blood vessel sprouting occurs in response to hypoxia. Consequently, vascular maturation is defective due to the sustained endothelial cell proliferation and migration. This results in leaky vascular architecture, with interrupted adherens junctions and inadequate blood flow (Figure 2). Indeed, hypoxia upregulates S1P<sub>2</sub> receptors in retinal endothelium and leads to formation of blood vessel sprouts with leaky basement membranes and limited perfusion.[48] In S1P<sub>2</sub> knockout mice, hypoxia-induced angiogenesis is characterized by formation of competent blood vessel sprouts that have comparable blood flow to the mature vasculature.[48] Similar results are seen with an S1P<sub>2</sub> receptor antagonist, as laser-induced choroidal neovascularization is

reduced after intravitreal injection of JTE013.[49] The effect of S1P<sub>2</sub> receptors on angiogenesis may vary in different endothelial cell types, as JTE013 administration increases S1P-mediated abdominal subcutaneous angiogenesis in mice.[78] In addition to angiogenic effects on retinal vasculature, S1P<sub>2</sub> activation with albumin-bound S1P results in disruption of barrier integrity and increased RPE permeability. Preincubation of RPE with JTE013 results in the repair of disrupted epithelium and reduced vascular leakage. Additionally, RPE shows increased VEGF release in response to S1P, an effect which is inhibited by JTE013, whereas the S1P<sub>1/3</sub> receptor antagonist VPC23019 has no effect.[49]

The net effect of S1P on vascular endothelium depends on the balance between S1P<sub>1</sub> and S1P<sub>2</sub> receptors. In vivo evidence show predominant expression of S1P<sub>2</sub> receptors in mouse CNV lesions,[49] upregulation of S1P<sub>2</sub> and S1P<sub>3</sub> receptors in light-induced damage in rat retinas,[33] and upregulation of S1P<sub>2</sub> receptors in a mouse model of retinal ischemia.[48] Additionally, S1P<sub>2</sub> knockout mice show enhanced retinal vascularization with normal vascular morphology following ischemic insult compared to *S1P<sub>2</sub><sup>+/+</sup>* mice[48]. Likewise intravitreal injection of S1P<sub>2</sub> antagonist JTE013 significantly reduced CNV lesion areas in mice.[49] This evidence explains why administration of anti-S1P antibodies reduces choroidal neovascularization and vessel leakage.[44,79] an action thought to be mediated by S1P<sub>2</sub> receptors. The type of S1P carrying molecules may also matter, as ApoM-bound S1P elicits a favorable activation of S1P<sub>1</sub> receptors resulting in reduced vascular leakage and increased expression of junction proteins in RPE, while albumin-bound S1P results in S1P<sub>2</sub>-mediated disruption of cellular junctions, increased vascular leakage and reduced endothelial integrity (Figure 2).[49]

### **Relationship between S1P and VEGF signaling**



VEGF affects S1P signaling in different ways. It increases S1P production by upregulating SphK expression in endothelial cells,[72] and also increases SphK activity in retinal endothelial cells (RECs).[80] While both actions raise intracellular S1P concentration, most of the intracellular S1P is transported to the extracellular space to act on S1P<sub>1-5</sub> receptors in autocrine and paracrine manners. VEGF specifically upregulates S1P<sub>1</sub> receptor expression in aortic endothelial cells, potentiating nitric oxide/Akt signaling, but has no effect on S1P<sub>2</sub> or S1P<sub>3</sub> receptor expression, suggesting that under hypoxic conditions, these endothelial cells might be more sensitive to S1P<sub>1</sub> signaling.[81] Yet, S1P<sub>1</sub> activation with SEW2871 blocks VEGF-induced sprouting in HUVECs and mouse microvascular endothelial cells, while the S1P<sub>1</sub> antagonist W146 increases VEGF-induced angiogenesis.[74] This suggests that upregulation of S1P<sub>1</sub> receptors in response to VEGF can lessen the overall angiogenic response to VEGF under pathological conditions. S1P results in increased VEGF expression in RPE cells, a response that is diminished after S1P<sub>2</sub> blockade with JTE013.[49] Additionally, S1P can transiently activate VEGFR2 receptors in bovine aortic endothelial cells in a tyrosine kinase inhibitor-sensitive manner, resulting in eNOS phosphorylation and activation.[82] SphK inhibition decreases VEGF-mediated RECs proliferation, migration and vascular leakage, indicating that S1P release is involved in VEGF-mediated angiogenesis.[80]

## **S1P ROLE IN OCULAR INFLAMMATION AND RELEASE OF INFLAMMATORY MEDIATORS**

Inflammatory processes also play a role in AMD, DR and RVO pathophysiology. Indeed, the lower incidence of retinopathy among diabetic patients on salicylate therapy for rheumatoid arthritis, and the significant effect of corticosteroid therapy on reducing macular edema and neovascularization in DR highlight the therapeutic relevance of anti-inflammatory drugs in

progression of DR.[83] Retinal ischemia is known to induce the expression of potent inflammatory cytokines including monocyte chemotactic protein-1 (MCP-1) and macrophage inflammatory protein-1 $\alpha$  (MIP-1 $\alpha$ ),[84] resulting in leukocyte infiltration and macrophage recruitment. Activated macrophage and microglia secrete inflammatory molecules such as TNF- $\alpha$  and interleukins,[85] which subsequently trigger a complex chain of cellular and vascular responses, the details of which are outside the scope of this review. Levels of MCP-1 are markedly increased in vitreous of patients with DR [86,87] and RVO.[87] Furthermore, markedly elevated IL-8 levels are detected in the vitreous fluid of DR and RVO patients,[87,88] higher IL-6 levels are also detected in the vitreous of DR patients,[86] and complement system activation is reported in AMD.[89] Which highlight the significant role of inflammatory response in neovascular ocular diseases.

In addition to its role in vascular integrity, S1P signaling can modulate inflammation. For instance, S1P reduces vascular leakage, neutrophils infiltration and lung edema after intratracheal administration of lipopolysaccharide.[90] But S1P also increases the production of inflammatory cytokines such as IL-8 and IL-6 among others.[43] S1P increases cyclooxygenase-2 (COX-2) expression and prostaglandin production via S1P<sub>2</sub> receptors in renal mesangial cells.[91] Fingolimod suppresses inflammation in an uveoretinitis model,[92] and inhibits leukocyte infiltration when administered as a single dose before induction of ocular inflammation.[93] Fingolimod-treated MS patients show a lower incidence of ocular inflammation compared to other MS patients.[94] It is unclear whether the ocular anti-inflammatory effects of fingolimod are due to its agonist or functional antagonist activity.[61,95] S1P increases IL-8, but not IL-6, production by RPE cells in a Pertussis toxin sensitive manner, suggesting S1P<sub>1</sub> receptor involvement.[43] However, another study suggests a role for S1P<sub>2</sub> receptors, as S1P-induced production of IL-8 and

CCL2 in RPE cells is decreased by JTE013, but not by S1P<sub>1</sub> or S1P<sub>3</sub> antagonists.[49] These apparently contradictory reports suggest that further work is needed to assess the role of different S1P receptors in retinal inflammation.

## **S1P ROLE IN PHOTORECEPTORS APOPTOSIS AND NEURODEGENERATION**

Retina, being a part of the CNS, comprises full neuronal circuits to acquire, convert and transfer electrical activity of photoreceptors to the brain, which is known as neural retina. Neural retina is a multilayered interconnected structure composed of five cell types, these are photoreceptors, bipolar cells, ganglion cells, horizontal cells, and amacrine cells.[96] Neurodegeneration in retinal diseases usually refer to apoptosis of retinal ganglionic cells and photoreceptors which leads to significant and progressive loss in visual function [24]. Retinal neurodegeneration is evident in diabetic retinopathy,[15] and neovascular AMD,[21] although the exact mechanism of neurodegeneration is not fully elucidated. Nevertheless, hypoxia-associated distorted retinal blood flow in these diseases is suggested to be the main reason to trigger neuronal death.[24] The role of S1P signaling in normal development of CNS is thoroughly reported, as sphingosine kinase null mice embryos suffered from neuronal tube defects with massive apoptosis in neuroepithelium.[97] Additionally, S1P signaling is involved in nerve growth factor mediated neurite extension,[98] and neuronal excitability.[99] S1P is required for development, differentiation and proliferation of photoreceptors in rat retinas.[51]

Under stress-induced photoreceptors and retinal ganglionic cells apoptosis, S1P can elicit different responses.[100,101] On one hand, S1P promotes cellular proliferation and reduces photoreceptor apoptosis.[102] Docosahexanoic acid (DHA, a mediator of photoreceptor survival and differentiation) increases intracellular S1P levels by upregulating SphK, and the protective

effects of DHA are blocked after SphK inhibition.[51] Similarly, action of S1P on S1P<sub>1</sub> receptors increases the survival of and mitigates the damage to retinal ganglionic cells following optic nerve injury,[103] and the selective S1P<sub>1</sub> receptor agonist CYM-5442 reduces retinal ganglionic cell damage after endothelin-1 induced vasoconstriction.[104] On the other hand, S1P acts as a pro-apoptotic mediator that can intensify the degenerative response in photoreceptors.[10] an action that is suggested to be mainly mediated by S1P<sub>2</sub> receptor activation.[35]

Under pathological conditions in the retina as hypoxic and oxidative stress, or optic nerve injury, S1P<sub>2</sub> receptors are upregulated while S1P<sub>1</sub> receptors are down-regulated.[33,103] This makes the role of S1P<sub>1</sub> receptors in ganglionic cell and photoreceptor survival is less obvious under pathological conditions.[103] At variance with the trophic effect of S1P<sub>1</sub> receptors, S1P/S1P<sub>2</sub> signaling elicits a detrimental effect on neuronal cells.[103]

## **S1P ROLE IN FIBROSIS, GLIOSIS AND EPITHELIAL TO MESENCHYMAL TRANSITION (EMT)**

Fibrosis is a reparative process that occurs in response to tissue injury, where the injured tissue is replaced by non-functional, collagen rich fibrous matrix. Outside the CNS, fibroblasts are the main players in fibrosis, as they migrate to the injured location, proliferate, synthesize and deposit extracellular matrix proteins.[105] As retina is considered a part of the CNS, the fibrotic response in the posterior chamber utilizes different mechanisms and cellular incorporation compared to that seen in non-CNS tissues.[106] Retina has a scarce fibroblast population; instead the fibrotic response is mainly mediated by RPE and Müller glial cells. RPE and glial cells are quiescent and non migratory under normal condition. Under inflammatory conditions or tissue injury, these cells undergo specific trans-differentiation to acquire fibroblast-like phenotype in processes known as EMT or gliosis. During EMT, RPE cells lose their epithelial traits and acquire

mesenchymal/fibroblast like phenotype, becoming invasive, migratory, lacking tight junction proteins and expressing mesenchymal markers.[107]. The fibrocontractile nature of transformed RPE can result in retinal detachment and severe vision impairment ending in further disease exacerbation. Similar transitional events occur in Müller cell gliosis, where Müller glial cells transdifferentiate to a fibroblast-like phenotype, release trophic mediators, and acquire proliferative and migratory properties.[108,109] Again, this transition to fibrocontractile structure results in gliotic scar tissue formation, which further exacerbates retinal damage.[110] While multiple cytokines and several proinflammatory signals can trigger a fibrotic response, there is growing evidence that TGF- $\beta$  is one of the most important cytokines that contribute to EMT [111,112] and gliosis.[113], as it is detected at higher levels in the vitreous of patients with DR.[114]

S1P signaling generally triggers fibrotic events in neovascular ocular disease. Administration of an anti-S1P antibody reduces collagen precipitation in sub-retinal structures after rupture of Bruch's membrane in a mouse model of CNV.[44] Likewise, locally injected anti-S1P monoclonal antibodies can alleviate conjunctival scarring following glaucoma filtering surgery.[115] S1P increases the production of contractile actin fibers and facilitates collagen deposition by RPE in vitro, one of the mesenchymal characteristics of EMT,[42] but the exact mechanism and S1P receptor subtype involved has not been reported. Migration of Müller glial cells in vitro is significantly increased by exogenously added S1P. Additionally, inhibition of SphK1, the main isoform in Müller glial cells, abolishes filopodia formation and cellular migration, suggesting that both endogenous and exogenous S1P amplify glial cell migration. S1P mediated actions are reduced by pre-treatment of Müller glial cells with the S1P<sub>3</sub> antagonist BML-241, suggesting that the effects are primarily mediated by S1P<sub>3</sub> receptors.[109]. Likewise, SphK1-

null mice show diminished gliosis and slower progression of Sandhoff disease, a central neurodegenerative disease. Similar results are obtained by S1P<sub>3</sub> receptor gene deletion.[116]

Although the relationship between S1P and TGF- $\beta$  signaling in neovascular ocular disease is not yet reported, there is established evidence of crosstalk between S1P and TGF- $\beta$  in renal mesangial cells.[117]. TGF- $\beta$  increases SphK1 in endometriotic stromal cells [118], human fibroblasts,[119] and in human kidney podocytes.[120] SphK1 upregulation in kidney is associated with protective rather than detrimental effects, as SphK1 deficient mice develop more drastic streptozocin induced nephropathy.[120]

## **CONCLUSION**

S1P is a promising therapeutic target that modulates angiogenesis, inflammation, apoptosis and fibrosis associated with neovascular ocular diseases. S1P/S1P<sub>1</sub> signaling can induce formation of competent blood vessel sprouts, increase retinal perfusion and reduce cell apoptosis and neurodegeneration. Blocking S1P<sub>2</sub> receptors achieves similar beneficial outcomes, while S1P<sub>3</sub> receptor antagonism or S1P<sub>1</sub> activation can inhibit gliosis. Under hypoxic conditions, SphK1 and S1P<sub>2</sub> are upregulated; this can be accompanied by S1P<sub>1</sub> downregulation, resulting in increased S1P production and predominant signaling through S1P<sub>2</sub> receptors. Therefore, inhibition of SphK1, S1P<sub>1</sub> activation and S1P<sub>2</sub>/S1P<sub>3</sub> antagonism might be used to attenuate retinal damage in ocular neovascular disease. Recent clinically approved S1P receptor modulators include siponimod and ozanimod; both can selectively activate S1P<sub>1</sub> receptors (along with S1P<sub>5</sub> receptors, which are less prevalent in ocular tissue). Evidence to date suggests that S1P receptor modulation plays important roles in the pathogenesis and treatment of neovascular ocular diseases. However, the role of these agents in the progression of neovascular ocular disease should be elucidated in preclinical models

to inform future clinical trials involving S1P receptor modulators already approved for other conditions.

## **ACKNOWLEDGMENTS**

Figure 1 was created in part using with BioRender. Elements of Figure 2 are from the Servier medical arts database.

## **COMPETING INTERESTS:**

NA

## **FUNDING**

This work was funded by the Irish research council (IRC).

## **CONTRIBUTORSHIP STATEMENT**

RAA, KBR, and CW contributed to the drafting of manuscript. RAA and CW contributed to the interpretation of the data in the literature. All authors contributed to the critical appraisal and final approval of the manuscript. CW provided the overall supervision of this work.

## REFERENCES

- 1 WHO. World report on vision. Geneva: World Health Organization; 2019.
- 2 Penn JS, Madan A, Caldwell RB, et al. Vascular endothelial growth factor in eye disease. *Prog Retin Eye Res* 2008;27:331–71.
- 3 Cunha-Vaz JG. The blood-retinal barriers. *Doc Ophthalmol* 1976;41:287–327.
- 4 Rizzolo LJ. RPE Polarity and Barrier Function. In: Klettner AK, Dithmar S, eds. *Retinal Pigment Epithelium in Health and Disease*. Berlin:Springer International Publishing 2020;19–45.
- 5 Gehrs KM, Anderson DH, Johnson LV, et al. Age-related macular degeneration-emerging pathogenetic and therapeutic concepts. *Annals of Medicine* 2006;38:450–71.
- 6 Ambati J, Fowler BJ. Mechanisms of age-related macular degeneration. *Neuron* 2012;75:26–39.
- 7 Campochiaro PA. Molecular Pathogenesis of Retinal and Choroidal Vascular Diseases. *Prog Retin Eye Res* 2015;49:67–81.
- 8 Aiello LP, Avery RL, Arrigg PG, et al. Vascular endothelial growth factor in ocular fluid of patients with diabetic retinopathy and other retinal disorders. *N Engl J Med* 1994;331:1480–7.
- 9 DeCroos FC, Todorich B, Alshareef R, et al. Neovascular Events in Eyes with Central Retinal Vein Occlusion Undergoing Serial Bevacizumab or Ranibizumab Intravitreal Injections: A Retrospective Review. *J Ophthalmic Vis Res* 2014;9:461–8.



- 10 Daruich A, Matet A, Moulin A, et al. Mechanisms of macular edema: Beyond the surface. *Prog Retin Eye Res* 2018;63:20–68.
- 11 Xu H-Z, Song Z, Fu S, et al. RPE barrier breakdown in diabetic retinopathy: seeing is believing. *J Ocul Biol Dis Infor* 2011;4:83–92.
- 12 Beatty S, Murray IJ, Henson DB, et al. Macular Pigment and Risk for Age-Related Macular Degeneration in Subjects from a Northern European Population. *Invest Ophthalmol Vis Sci* 2001;42:439–46.
- 13 Fisher CR, Ferrington DA. Perspective on AMD Pathobiology: A Bioenergetic Crisis in the RPE. *Invest Ophthalmol Vis Sci* 2018;59:AMD41–7.
- 14 Park S-H, Park J-W, Park S-J, et al. Apoptotic death of photoreceptors in the streptozotocin-induced diabetic rat retina. *Diabetologia* 2003;46:1260–8.
- 15 Kanamori A, Nakamura M, Mukuno H, et al. Diabetes has an additive effect on neural apoptosis in rat retina with chronically elevated intraocular pressure. *Curr Eye Res* 2004;28:47–54.
- 16 Portera-Cailliau C, Sung CH, Nathans J, et al. Apoptotic photoreceptor cell death in mouse models of retinitis pigmentosa. *Proc Natl Acad Sci USA* 1994;91:974–8.
- 17 Buschini E, Piras A, Nuzzi R, et al. Age related macular degeneration and drusen: Neuroinflammation in the retina. *Prog Neurobiol* 2011;95:14–25.
- 18 Meleth AD, Agrón E, Chan C-C, et al. Serum inflammatory markers in diabetic retinopathy. *Invest Ophthalmol Vis Sci* 2005;46:4295–301.

- 19 Whitcup SM, Nussenblatt RB, Lightman SL, et al. Inflammation in retinal disease. *Int J Inflam* 2013;2013:724648.
- 20 Rivera JC, Dabouz R, Noueihed B, et al. Ischemic Retinopathies: Oxidative Stress and Inflammation. *Oxid Med Cell Longev* 2017;2017:3940241.
- 21 Medeiros NE, Curcio CA. Preservation of ganglion cell layer neurons in age-related macular degeneration. *Invest Ophthalmol Vis Sci* 2001;42:795–803.
- 22 Zafar S, Sachdeva M, Frankfort BJ, et al. Retinal Neurodegeneration as an Early Manifestation of Diabetic Eye Disease and Potential Neuroprotective Therapies. *Curr Diab Rep* 2019;19:17.
- 23 Almasieh M, Wilson AM, Morquette B, et al. The molecular basis of retinal ganglion cell death in glaucoma. *Prog Retin Eye Res* 2012;31:152–81.
- 24 Osborne NN, Chidlow G, Layton CJ, et al. Optic nerve and neuroprotection strategies. *Eye (Lond)* 2004;18:1075–84.
- 25 Little K, Ma JH, Yang N, et al. Myofibroblasts in macular fibrosis secondary to neovascular age-related macular degeneration - the potential sources and molecular cues for their recruitment and activation. *EBioMedicine* 2018;38:283–91.
- 26 Roy S, Amin S, Roy S. Retinal Fibrosis in Diabetic Retinopathy. *Exp Eye Res* 2016;142:71–5.
- 27 Ghasemi Falavarjani K, Nguyen QD. Adverse events and complications associated with intravitreal injection of anti-VEGF agents: a review of literature. *Eye* 2013;27:787–94.

- 28 Young MM, Kester M, Wang H-G. Sphingolipids: regulators of crosstalk between apoptosis and autophagy. *J Lipid Res* 2013;54:5–19.
- 29 Gault C, Obeid L, Hannun Y. An overview of sphingolipid metabolism: from synthesis to breakdown. *Adv Exp Med Biol* 2010;688:1–23.
- 30 Maceyka M, Sankala H, Hait NC, et al. SphK1 and SphK2, sphingosine kinase isoenzymes with opposing functions in sphingolipid metabolism. *J Biol Chem* 2005;280:37118–29.
- 31 Spiegel S, Milstien S. Sphingosine-1-phosphate: an enigmatic signalling lipid. *Nat Rev Mol Cell Biol* 2003;4:397–407.
- 32 Proia RL, Hla T. Emerging biology of sphingosine-1-phosphate: its role in pathogenesis and therapy. *J Clin Invest* 2015;125:1379–87.
- 33 Porter H, Qi H, Prabhu N, et al. Characterizing Sphingosine Kinases and Sphingosine 1-Phosphate Receptors in the Mammalian Eye and Retina. *Int J Mol Sci* 2018;19:3885.
- 34 Eresch J, Stumpf M, Koch A, et al. Sphingosine Kinase 2 Modulates Retinal Neovascularization in the Mouse Model of Oxygen-Induced Retinopathy. *Invest Ophthalmol Vis Sci* 2018;59:653–61.
- 35 Terao R, Honjo M, Ueta T, et al. Light Stress-Induced Increase of Sphingosine 1-Phosphate in Photoreceptors and Its Relevance to Retinal Degeneration. *Int J Mol Sci* 2019;20.
- 36 Panova IG, Tatikolov AS, Smirnova YuA, et al. Albumin in the Vitreous Body, Retina and Lens of Human Fetal Eye. *Bull Exp Biol Med* 2017;162:629–31.

- 37 Pikuleva IA, Curcio CA. Cholesterol in the retina: the best is yet to come. *Prog Retin Eye Res* 2014;0:64–89.
- 38 Tserentsoodol N, Sztejn J, Campos M, et al. Uptake of cholesterol by the retina occurs primarily via a low density lipoprotein receptor-mediated process. *Mol Vis* 2006;12:1306–18.
- 39 Amaratunga A, Abraham CR, Edwards RB, et al. Apolipoprotein E Is Synthesized in the Retina by Müller Glial Cells, Secreted into the Vitreous, and Rapidly Transported into the Optic Nerve by Retinal Ganglion Cells. *J Biol Chem* 1996;271:5628–32.
- 40 Kluk MJ, Hla T. Signaling of sphingosine-1-phosphate via the S1P/EDG-family of G-protein-coupled receptors. *Biochimica et Biophysica Acta (BBA) - Molecular and Cell Biology of Lipids* 2002;1582:72–80.
- 41 Kono M, Mi Y, Liu Y, et al. The sphingosine-1-phosphate receptors S1P1, S1P2, and S1P3 function coordinately during embryonic angiogenesis. *J Biol Chem* 2004;279:29367–73.
- 42 Swaney JS, Moreno KM, Gentile AM, et al. Sphingosine-1-phosphate (S1P) is a novel fibrotic mediator in the eye. *Exp Eye Res* 2008;87:367–75.
- 43 Qiao Y, Hu R, Wang Q, et al. Sphingosine 1-Phosphate Elicits Proinflammatory Responses in ARPE-19 Cells. *Invest Ophthalmol Vis Sci* 2012;53:8200–7.
- 44 Caballero S, Swaney J, Moreno K, et al. Anti-sphingosine-1-phosphate monoclonal antibodies inhibit angiogenesis and sub-retinal fibrosis in a murine model of laser-induced choroidal neovascularization. *Exp Eye Res* 2009;88:367–77.

- 45 Menon M, Mohammadi S, Davila-Velderrain J, et al. Single-cell transcriptomic atlas of the human retina identifies cell types associated with age-related macular degeneration. *Nat Commun* 2019;10:4902.
- 46 Voigt AP, Mulfaul K, Mullin NK, et al. Single-cell transcriptomics of the human retinal pigment epithelium and choroid in health and macular degeneration. *Proc Natl Acad Sci USA* 2019;116:24100–7.
- 47 Voigt AP, Binkley E, Flamme-Wiese MJ, et al. Single-Cell RNA Sequencing in Human Retinal Degeneration Reveals Distinct Glial Cell Populations. *Cells* 2020;9:438.
- 48 Skoura A, Sanchez T, Claffey K, et al. Essential role of sphingosine 1–phosphate receptor 2 in pathological angiogenesis of the mouse retina. *J Clin Invest* 2007;117:2506–16.
- 49 Terao R, Honjo M, Totsuka K, et al. The role of sphingosine 1-phosphate receptors on retinal pigment epithelial cells barrier function and angiogenic effects. *Prostaglandins & Other Lipid Mediat* 2019;145:106365.
- 50 Yanagida K, Engelbrecht E, Niaudet C, et al. Sphingosine 1-Phosphate Receptor Signaling Establishes AP-1 Gradients to Allow for Retinal Endothelial Cell Specialization. *Dev Cell* 2020;52:779-793.
- 51 Miranda GE, Abrahan CE, Politi LE, et al. Sphingosine-1-Phosphate Is a Key Regulator of Proliferation and Differentiation in Retina Photoreceptors. *Invest Ophthalmol Vis Sci* 2009;50:4416.

- 52 Chen H, Chan AY, Stone DU, et al. Beyond the cherry-red spot: Ocular manifestations of sphingolipid-mediated neurodegenerative and inflammatory disorders. *Surv Ophthalmol* 2014;59:64–76.
- 53 Simón MV, Prado Spalm FH, Vera MS, et al. Sphingolipids as Emerging Mediators in Retina Degeneration. *Front Cell Neurosci* 2019;13:246.
- 54 Penno A, Reilly MM, Houlden H, et al. Hereditary Sensory Neuropathy Type 1 Is Caused by the Accumulation of Two Neurotoxic Sphingolipids. *J Biol Chem* 2010;285:11178–87.
- 55 Gantner ML, Eade K, Wallace M, et al. Serine and Lipid Metabolism in Macular Disease and Peripheral Neuropathy. *N Engl J Med* 2019;381:1422–33.
- 56 Simon MV, Basu SK, Qaladize B, et al. Sphingolipids as critical players in retinal physiology and pathology. *J Lipid Res* 2021;62:100037.
- 57 Terao R, Honjo M, Aihara M. Apolipoprotein M Inhibits Angiogenic and Inflammatory Response by Sphingosine 1-Phosphate on Retinal Pigment Epithelium Cells. *Int J Mol Sci* 2017;19:112.
- 58 Salomone S, Waeber C. Selectivity and Specificity of Sphingosine-1-Phosphate Receptor Ligands: Caveats and Critical Thinking in Characterizing Receptor-Mediated Effects. *Front Pharmacol* 2011;2:9.
- 59 Derfuss T, Mehling M, Papadopoulou A, et al. Advances in oral immunomodulating therapies in relapsing multiple sclerosis. *Lancet Neurol* 2020;19:336–47.

- 60 Chew WS, Wang W, Herr DR. To fingolimod and beyond: The rich pipeline of drug candidates that target S1P signaling. *Pharmacol Res* 2016;113:521–32.
- 61 Bigaud M, Guerini D, Billich A, et al. Second generation S1P pathway modulators: Research strategies and clinical developments. *Biochim Biophys Acta* 2014;1841:745–58.
- 62 Jain N, Bhatti MT. Fingolimod-associated macular edema: incidence, detection, and management. *Neurology* 2012;78:672–80.
- 63 Nørgaard TL, Andersen CU, Hilt C, et al. Macular oedema and changes in macular thickness in multiple sclerosis patients treated with fingolimod. *Basic Clin Pharmacol Toxicol* 2020;126:492–7.
- 64 Ribatti D. Angiogenesis. In: Maloy S, Hughes K, eds. *Brenner’s Encyclopedia of Genetics* (Second Edition). San Diego:Academic Press 2013;30–132.
- 65 Warmke N, Walker AMN, Cubbon RM. Angiogenesis. In: Vasan RS, Sawyer DB, eds. *Encyclopedia of Cardiovascular Research and Medicine*. Oxford:Elsevier 2018;85–96.
- 66 Jung B, Obinata H, Galvani S, et al. Flow-Regulated Endothelial S1P Receptor-1 Signaling Sustains Vascular Development. *Dev Cell* 2012;23:600–10.
- 67 Qazi Y, Maddula S, Ambati BK. Mediators of ocular angiogenesis. *J Genet* 2009;88:495–515.
- 68 Gerritsen ME. Angiogenesis. In: Tuma RF, Durán WN, Ley K, eds. *Microcirculation* (Second Edition). San Diego:Academic Press 2008;351–383.

- 69 Cuvillier O, Ader I, Bouquerel P, *et al.* Hypoxia, Therapeutic Resistance, and Sphingosine 1-Phosphate. In: Norris JS, ed. *Advances in Cancer Research*. San Diego:Academic Press 2013;117–141.
- 70 Oyama O, Sugimoto N, Qi X, *et al.* The lysophospholipid mediator sphingosine-1-phosphate promotes angiogenesis in vivo in ischaemic hindlimbs of mice. *Cardiovasc Res* 2008;78:301–7. doi:10.1093/cvr/cvn002
- 71 Obinata H, Kuo A, Wada Y, *et al.* Identification of ApoA4 as a sphingosine 1-phosphate chaperone in ApoM- and albumin-deficient mice. *J Lipid Res* 2019;60:1912–21.
- 72 Brinkmann V. Sphingosine 1-phosphate receptors in health and disease: Mechanistic insights from gene deletion studies and reverse pharmacology. *Pharmacol Ther* 2007;115:84–105.
- 73 Waeber C. Sphingosine 1-Phosphate (S1P) Signaling and the Vasculature. In: Chun J, Hla T, Spiegel S, *et al.*, eds. *Lysophospholipid Receptors*. Hoboken, NJ, USA:John Wiley & Sons Inc.2013;313–47.
- 74 Gaengel K, Niaudet C, Hagikura K, *et al.* The Sphingosine-1-Phosphate Receptor S1PR1 Restricts Sprouting Angiogenesis by Regulating the Interplay between VE-Cadherin and VEGFR2. *Dev Cell* 2012;23:587–99.
- 75 Cartier A, Leigh T, Liu CH, *et al.* Endothelial sphingosine 1-phosphate receptors promote vascular normalization and antitumor therapy. *Proc Natl Acad Sci USA* 2020;117:3157–66.



- 76 Lee J-F, Gordon S, Estrada R, et al. Balance of S1P1 and S1P2 signaling regulates peripheral microvascular permeability in rat cremaster muscle vasculature. *Am J Physiol Heart Circ Physiol* 2009;296:H33–42.
- 77 Sanchez T, Skoura A, Wu MT, et al. Induction of vascular permeability by the sphingosine-1-phosphate receptor-2 (S1P2R) and its downstream effectors ROCK and PTEN. *Arterioscler Thromb Vasc Biol* 2007;27:1312–8.
- 78 Inoki I, Takuwa N, Sugimoto N, *et al.* Negative regulation of endothelial morphogenesis and angiogenesis by S1P2 receptor. *Biochem Biophys Res Commun* 2006;346:293–300.
- 79 Xie B, Shen J, Dong A, et al. Blockade of Sphingosine-1-phosphate Reduces Macrophage Influx and Retinal and Choroidal Neovascularization. *J Cell Physiol* 2009;218:192–8.
- 80 Maines LW, French KJ, Wolpert EB, et al. Pharmacologic Manipulation of Sphingosine Kinase in Retinal Endothelial Cells: Implications for Angiogenic Ocular Diseases. *Invest Ophthalmol Vis Sci* 2006;47:5022.
- 81 Igarashi J, Erwin PA, Dantas APV, et al. VEGF induces S1P1 receptors in endothelial cells: Implications for cross-talk between sphingolipid and growth factor receptors. *Proc Natl Acad Sci U S A* 2003;100:10664–9.
- 82 Tanimoto T, Jin Z-G, Berk BC. Transactivation of vascular endothelial growth factor (VEGF) receptor Flk-1/KDR is involved in sphingosine 1-phosphate-stimulated phosphorylation of Akt and endothelial nitric-oxide synthase (eNOS). *J Biol Chem* 2002;277:42997–3001.

- 83 Rezzola S, Loda A, Corsini M, et al. Angiogenesis-Inflammation Cross Talk in Diabetic Retinopathy: Novel Insights From the Chick Embryo Chorioallantoic Membrane/Human Vitreous Platform. *Front Immunol* 2020;11:581288
- 84 Yoshida S, Yoshida A, Ishibashi T, et al. Role of MCP-1 and MIP-1 $\alpha$  in retinal neovascularization during postischemic inflammation in a mouse model of retinal neovascularization. *J Leukoc Biol* 2003;73:137-44.
- 85 Dell’Omo R, Semeraro F, Bamonte G, et al. Vitreous Mediators in Retinal Hypoxic Diseases. *Mediators Inflamm* 2013;2013:935301.
- 86 Yoshida S, Kubo Y, Kobayashi Y, et al. Increased vitreous concentrations of MCP-1 and IL-6 after vitrectomy in patients with proliferative diabetic retinopathy: possible association with postoperative macular oedema. *Br J Ophthalmol* 2015;99:960–6.
- 87 Yoshimura T, Sonoda K-H, Sugahara M, et al. Comprehensive Analysis of Inflammatory Immune Mediators in Vitreoretinal Diseases. *PLoS One* 2009;4:e8158.
- 88 Dan-Brezi I, Zahavi A, Axer-Siegel R, et al. Inflammation, angiogenesis and coagulation interplay in a variety of retinal diseases. *Acta Ophthalmol* 2020;98:e559-e562
- 89 Donoso LA, Kim D, Frost A, et al. The Role of Inflammation in the Pathogenesis of Age-related Macular Degeneration. *Surv Ophthalmol* 2006;51:137–52.
- 90 Peng X, Hassoun PM, Sammani S, et al. Protective Effects of Sphingosine 1-Phosphate in Murine Endotoxin-induced Inflammatory Lung Injury. *Am J Respir Crit Care Med* 2004;169:1245–51.

- 91 Völzke A, Koch A, Meyer Zu Heringdorf D, et al. Sphingosine 1-phosphate (S1P) induces COX-2 expression and PGE2 formation via S1P receptor 2 in renal mesangial cells. *Biochim Biophys Acta* 2014;1841:11–21.
- 92 Commodaro AG, Peron JPS, Lopes CT, et al. Evaluation of experimental autoimmune uveitis in mice treated with FTY720. *Invest Ophthalmol Vis Sci* 2010;51:2568–74.
- 93 Raveney BJE, Copland DA, Nicholson LB, et al. Fingolimod (FTY720) as an acute rescue therapy for intraocular inflammatory disease. *Arch Ophthalmol* 2008;126:1390–5.
- 94 Sonne SJ, Smith BT. Incidence of uveitis and macular edema among patients taking fingolimod 0.5 mg for multiple sclerosis. *J Ophthalmic Inflamm Infect* 2020;10:24.
- 95 Sanna MG, Wang S-K, Gonzalez-Cabrera PJ, et al. Enhancement of capillary leakage and restoration of lymphocyte egress by a chiral S1P1 antagonist in vivo. *Nat Chem Biol* 2006;2:434–41. doi:10.1038/nchembio804
- 96 Neuroscience. 2nd edition. Purves D, Augustine GJ, Fitzpatrick D, et al., eds. Sunderland (MA):Sinauer Associates 2001.
- 97 Mizugishi K, Yamashita T, Olivera A, et al. Essential Role for Sphingosine Kinases in Neural and Vascular Development. *Mol Cell Biol*.2005;25:11113-21
- 98 Toman RE, Payne SG, Watterson KR, et al. Differential transactivation of sphingosine-1-phosphate receptors modulates NGF-induced neurite extension. *J Cell Biol* 2004;166:381-92.

- 99 MacLennan AJ, Carney PR, Zhu WJ, et al. An essential role for the H218/AGR16/Edg-5/LP(B2) sphingosine 1-phosphate receptor in neuronal excitability. *Eur J Neurosci* 2001;14:203–9.
- 100 Acharya U. Modulating Sphingolipid Biosynthetic Pathway Rescues Photoreceptor Degeneration. *Science* 2003;299:1740–3.
- 101 Sugano E, Edwards G, Saha S, et al. Overexpression of acid ceramidase (ASAH1) protects retinal cells (ARPE19) from oxidative stress. *Journal of Lipid Research* 2019;60:30–43.
- 102 Abrahan CE, Miranda GE, Agnolazza DL, et al. Synthesis of Sphingosine Is Essential for Oxidative Stress-Induced Apoptosis of Photoreceptors. *Invest Ophthalmol Vis Sci* 2010;51:1171-80.
- 103 Joly S, Pernet V. Sphingosine 1-phosphate receptor 1 is required for retinal ganglion cell survival after optic nerve trauma. *J Neurochem* 2016;138:571–86.
- 104 Blanco R, Martínez-Navarrete G, Valiente-Soriano FJ, et al. The S1P1 receptor-selective agonist CYM-5442 protects retinal ganglion cells in endothelin-1 induced retinal ganglion cell loss. *Exp Eye Res* 2017;164:37–45.
- 105 Kendall RT, Feghali-Bostwick CA. Fibroblasts in fibrosis: novel roles and mediators. *Front Pharmacol* 2014;5:123.
- 106 Friedlander M. Fibrosis and diseases of the eye. *J Clin Invest* 2007;117:576–86.
- 107 Zhou M, Geathers JS, Grillo SL, et al. Role of Epithelial-Mesenchymal Transition in Retinal Pigment Epithelium Dysfunction. *Front Cell Dev Biol* 2020;8:501.

- 108 Abrahan CE, Insua MF, Politi LE, et al. Oxidative stress promotes proliferation and dedifferentiation of retina glial cells in vitro. *J Neurosci Res* 2009;87:964–77.
- 109 Simón MV, Spalm FHP, Politi LE, et al. Sphingosine-1-Phosphate Is a Crucial Signal for Migration of Retina Müller Glial Cells. *Invest Ophthalmol Vis Sci* 2015;56:5808–15.
- 110 Guidry C. The role of Müller cells in fibrocontractive retinal disorders. *Prog Retin Eye Res* 2005;24:75-86
- 111 Kobayashi M, Tokuda K, Kobayashi Y, et al. Suppression of Epithelial-Mesenchymal Transition in Retinal Pigment Epithelial Cells by an MRTF-A Inhibitor. *Invest Ophthalmol Vis Sci* 2019;60:528–37.
- 112 Xiao W, Chen X, Liu X, et al. Trichostatin A, a histone deacetylase inhibitor, suppresses proliferation and epithelial–mesenchymal transition in retinal pigment epithelium cells. *J Cell Mol Med* 2014;18:646–55.
- 113 Fan J, Shen W, Lee S-R, et al. Targeting the Notch and TGF- $\beta$  signaling pathways to prevent retinal fibrosis in vitro and in vivo. *Theranostics* 2020;10:7956–73.
- 114 Kita T, Hata Y, Kano K, et al. Transforming Growth Factor- $\beta$ 2 and Connective Tissue Growth Factor in Proliferative Vitreoretinal Diseases: Possible Involvement of Hyalocytes and Therapeutic Potential of Rho Kinase Inhibitor. *Diabetes* 2007;56:231–8.
- 115 Lukowski ZL, Min J, Beattie AR, et al. Prevention of ocular scarring after glaucoma filtering surgery using the monoclonal antibody LT1009 (Sonpeizumab) in a rabbit model. *J Glaucoma* 2013;22:145–51.

- 116 Wu Y-P, Mizugishi K, Bektas M, et al. Sphingosine kinase 1/S1P receptor signaling axis controls glial proliferation in mice with Sandhoff disease. *Hum Mol Genet* 2008;17:2257–64.
- 117 Xin C, Ren S, Kleuser B, et al. Sphingosine 1-phosphate cross-activates the Smad signaling cascade and mimics transforming growth factor-beta-induced cell responses. *J Biol Chem* 2004;279:35255–62.
- 118 Yoshino O, Yamada-Nomoto K, Kano K, et al. Sphingosine I Phosphate (S1P) Increased IL-6 Expression and Cell Growth in Endometriotic Cells. *Reprod Sci* 2019;26:1460–7.
- 119 Yamanaka M, Shegogue D, Pei H, et al. Sphingosine Kinase 1 (SPHK1) Is Induced by Transforming Growth Factor- $\beta$  and Mediates TIMP-1 Up-regulation. *J Biol Chem* 2004;279:53994–54001.
- 120 Ren S, Babelova A, Moreth K, et al. Transforming growth factor- $\beta$ 2 upregulates sphingosine kinase-1 activity, which in turn attenuates the fibrotic response to TGF- $\beta$ 2 by impeding CTGF expression. *Kidney Int* 2009;76:857–67.

## FIGURE LEGEND

### Figure 1.

Schematic representation of synthesis and metabolism of sphingolipids, with special emphasis on S1P and the 5 receptor subtypes it can activate (S1P<sub>1-5</sub> receptors) as well as the G proteins they are coupled to. Specific S1P receptor modulators discussed in this review are listed at the bottom of the figure (blue and red circles indicate agonist and antagonist activities, respectively). This figure was created by the authors, using the software Biorender.

### Figure 2.

Summary of S1P-mediated effects on angiogenesis. Top: S1P<sub>1</sub> receptor activation by S1P reduces proangiogenic factors release in response to hypoxia (most importantly VEGF) leading to fewer tip cell formation and fewer branching points per unit area. This leads to formation of fewer vessel branches. S1P<sub>1</sub> receptors also increase intercellular junction proteins expression and perivascular cells coverage of newly formed sprouts. This leads to formation of competent blood vessels with normal blood flow that restore tissue perfusion (red areas) and downregulate the angiogenic signal. Bottom: Hypoxia results in over-expression of SphK1 and S1P<sub>2</sub> receptors. This receptor subtype increases VEGF release and tip cells number resulting in increased vessel branching per unit area. The formed branches have defective expression of intercellular junction proteins and irregular perivascular cells coverage. This leads to formation of leaky endothelium with diminished blood flow which further exacerbate tissue hypoxia (blue areas). Due to sustained angiogenesis, further branching of the new sprouts occurs leading to further leaking and hemorrhage. The net effect of these two opposite signals depends on relative receptors densities and specific receptor upregulation in response to hypoxia. Although their abundance in the eye is not known, the balance

of S1P carrier proteins may also play a role, as ApoM-bound S1P and Albumin-bound S1P preferentially activate S1P<sub>1</sub> and S1P<sub>2</sub> mediated cascades, respectively. This figure was created by the authors using elements from the Servier medical arts database.



Figure 1:

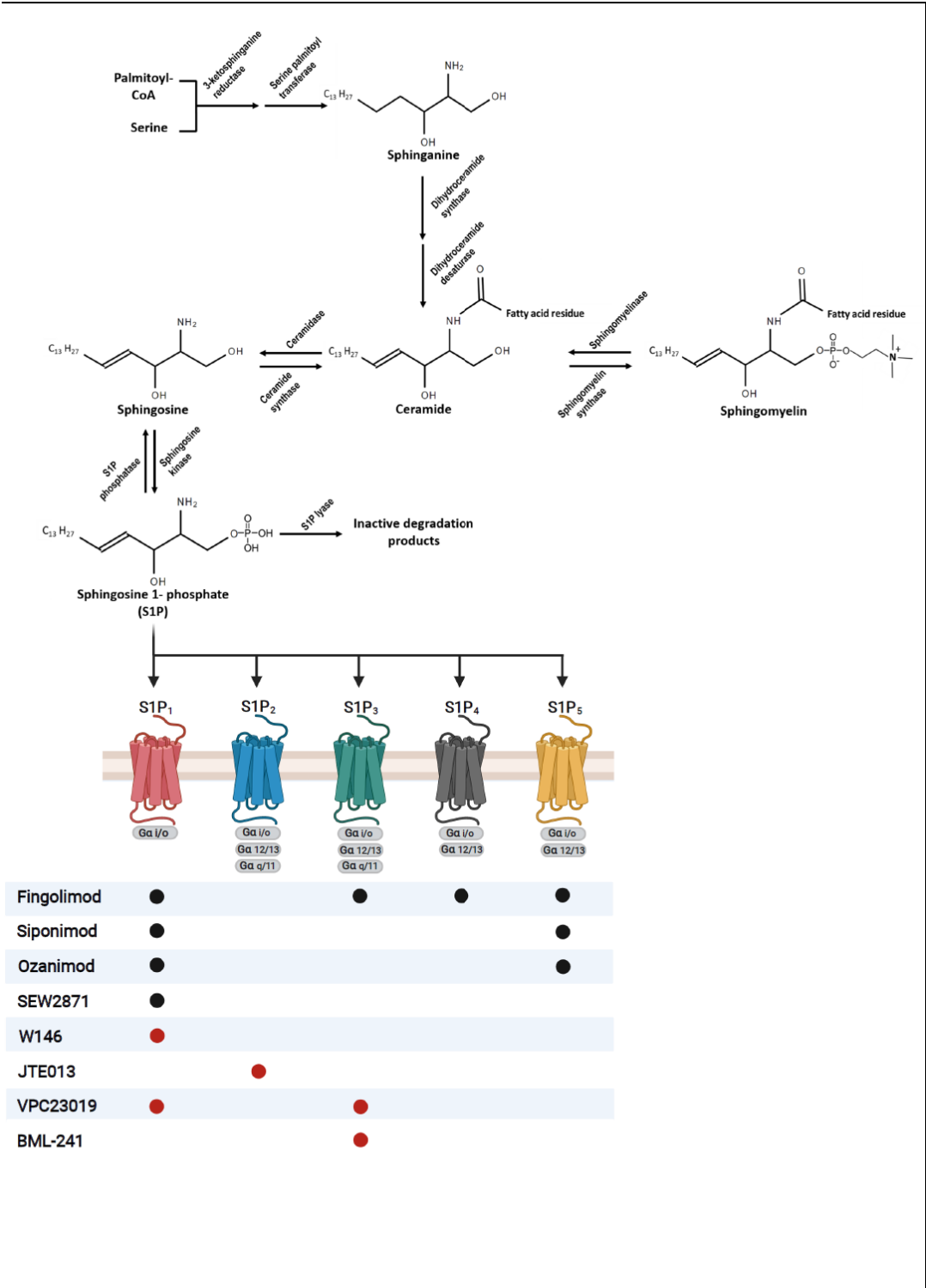


Figure 2

